

Photochemical efficiency and variation in photosynthetic pigments in *Nothofagus pumilio* seedlings growing under light intensity and soil moisture gradients

Lucía Bottan ⁽¹⁾,
María Vanessa Lencinas ⁽¹⁾,
Pablo Luis Peri ⁽²⁾,
Miriam Arena ⁽³⁾,
Guillermo Martínez Pastur ⁽¹⁾

Regeneration dynamics in forest ecosystems depend on seedlings' ability to persist in shaded understories and, following canopy opening, to rapidly adjust physiologically to new, contrasting light and soil moisture conditions. In this study, we examined the real-time efficiency of the photosynthetic apparatus and the short- to intermediate-term acclimatory response of photosynthetic pigments on *Nothofagus pumilio* seedlings exposed to different light and soil moisture levels. We evaluated phenology, chlorophyll fluorescence and pigment content in leaves, on 2-3 years-old seedlings in a controlled greenhouse experiment with three levels of light intensity (low = 4%, medium = 26%, high = 64% of the natural incident irradiance) and two levels of soil moisture (optimal = 40-60%, excess = 80-100% of soil field capacity). We measured six samples per treatment (light intensity × soil moisture levels) monthly during one growing season (n = 216). Data were analyzed by multiple ANOVAs. Although maximum quantum yield remained stable (> 0.80) across treatments, seedlings exhibited differential eco-physiological responses primarily driven by light availability. Structural investment was synergistic and negatively affected by the light × soil moisture interaction, resulting in reduced leaf production, chlorophyll, and carotenoids during peak season under the high-light and excess-soil-moisture treatment. Seedlings under medium light showed optimal performance, with maximum pigment accumulation and high PSII efficiency without signs of chronic photoinhibition. *N. pumilio* seedlings decouple short-term functional photochemical efficiency from their structural investment strategy (pigment content), a mechanism that ensures survival under variable canopy cover but results in reduced growth under sub-optimal conditions.

□ (1) Laboratorio de Recursos Agroforestales, Centro Austral de Investigaciones Científicas - CADIC, Consejo Nacional de Investigaciones Científicas y Técnicas - CONICET, cc 92, 9410 Ushuaia, Tierra del Fuego (Argentina); (2) Instituto Nacional de Tecnología Agropecuaria - INTA, Universidad Nacional de La Patagonia Austral - UNPA, Consejo Nacional de Investigaciones Científicas y Técnicas - CONICET, cc 332, 9400, Río Gallegos, Santa Cruz (Argentina); (3) CONICET - Laboratorio de Fisiología Vegetal, Instituto de Ciencias de la Vida, SeCyT - Rectorado, Universidad de Morón, Machado 914, B1708EOH, Morón, Buenos Aires (Argentina)

@ Lucía Bottan (luciaabottan@gmail.com)

Received: Oct 24, 2025 - Accepted: Jun 22, 2026

Citation: Bottan L, Lencinas MV, Peri PL, Arena M, Martínez Pastur G (2026). Photochemical efficiency and variation in photosynthetic pigments in *Nothofagus pumilio* seedlings growing under light intensity and soil moisture gradients. iForest 19: 311-320. - doi: [10.3832/ifor5020-019](https://doi.org/10.3832/ifor5020-019) [online 2026-08-19]

Communicated by: Luigi Saulino

Keywords: Acclimation, Chlorophyll Fluorescence, Photoprotection, Phenotypic Plasticity, *Nothofagus pumilio*

Introduction

Understanding forest regeneration dynamics is essential for ensuring their conservation and predicting their responses to natural and anthropogenic disturbances. *Nothofagus pumilio* (Poepp. et Endl.) Krasser is a native tree species of high ecological and economic importance, widely distributed across Chilean and Argentine Patagonia (Donoso et al. 2022). In these forests, tree regeneration commonly persists in the shaded understory for extended periods (Rebertus & Veblen 1993, Rodríguez-Souilla et al. 2023). This leads to the formation of a seedling bank (Cuevas & Arroyo 1999) that responds to canopy gaps caused by windstorms, logging, exotic beaver activity, or avalanches (Heinemann et al. 2000, Henn et al. 2016, Martínez Pastur et al. 2024). As regeneration in southern temperate forests is largely driven by gap dynamics, current silvicultural prescriptions for *N. pumilio* forests in Tierra del Fuego focus on creating canopy openings to promote natural regeneration (Chaves et al. 2024). These management approaches are implemented through harvesting systems such as shelterwood cuts (Martínez Pastur et al. 2000)

and variable-retention harvesting (Gustafsson et al. 2012), which modify canopy structure and resource availability. Light and soil moisture are among the most critical factors affecting the growth and survival of understory seedlings in austral forests (Heinemann & Kitzberger 2006, Martínez Pastur et al. 2011a, Rodríguez-Souilla et al. 2023). As a mid-tolerant to shade species, *N. pumilio* responds positively to moderate light intensity (Martínez Pastur et al. 2011b). Although low soil water availability is a known limiting factor, excessive soil moisture can also negatively affect seedling performance, particularly in mesic forest species (Herrera-Bevan & Ibáñez 2024). For example, in *N. pumilio* forests, site quality decreases when soil moisture exceeds around 60% of field capacity (Martínez Pastur et al. 2007), as reduced soil aeration and altered nutrient dynamics constrain root functioning and resource uptake (Peri et al. 2009, Zhang et al. 2025). Although several studies evaluated *N. pumilio* seedlings under extreme conditions, the eco-physiological mechanisms underlying their responses to sudden changes in light and soil resources remain poorly understood

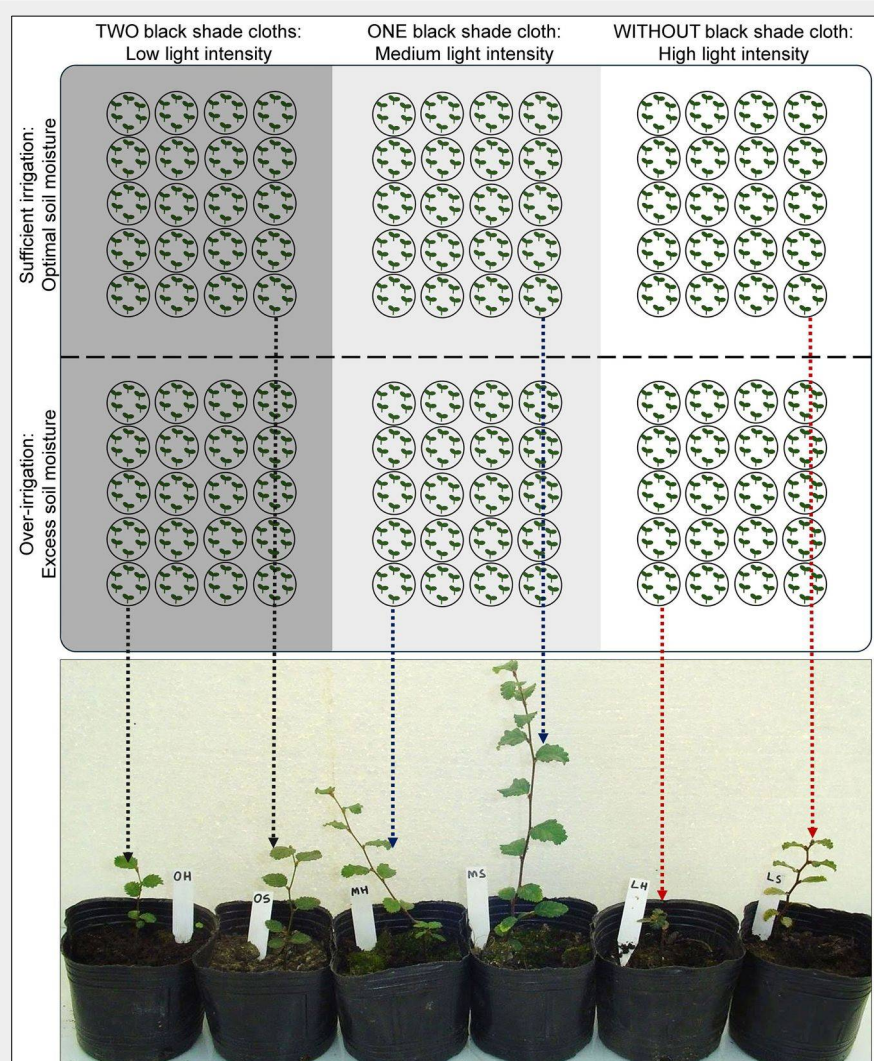


Fig. 1 - Conceptual diagram of the arrangement of *Nothofagus pumilio* seedlings within the greenhouse, along with a comparative photo of seedlings for each treatment taken at the end of the growing season (March). Six blocks were established for each combination of soil moisture treatments (optimal soil moisture; excess soil moisture) and light intensity treatments (low light intensity; medium light intensity; high light intensity).

impairment under environmental stress. In this context, evaluating physiological traits can provide valuable insights into stress levels, light-use efficiency, and the growth potential of tree seedlings under different understory conditions.

To analyze the effects of light and soil moisture on *N. pumilio* seedling physiology, we conducted a controlled greenhouse experiment. This setup simulated different harvesting scenarios, such as those resulting from canopy openings created by variable retention and shelterwood cuts in Tierra del Fuego (Martínez Pastur et al. 2007, 2011a, 2011b, Ivancich et al. 2012). The objective was to estimate the real-time efficiency of the photosynthetic apparatus and the short- to intermediate-term acclimatory response of photosynthetic pigments in *N. pumilio* seedlings exposed to different light intensities and soil moisture levels. We assessed this by measuring chlorophyll fluorescence and photosynthetic pigment content in leaves over a six-month growing season. We hypothesized that light intensity and soil moisture interact synergistically to determine the physiological performance of *N. pumilio* seedlings, such that extreme combinations (high light and excess soil moisture) reduce photochemical efficiency (Φ_{PSII} , qP) and increase photoprotective responses (NPQ and carotenoids). In contrast, intermediate light conditions and optimal soil moisture promote higher chlorophyll content and maintain photochemical performance, resulting in a unimodal physiological response across the light gradient modulated by soil moisture availability. This approach allows understanding of how *N. pumilio* natural regeneration responds to contrasting environments in terms of photosynthetic function and acclimation to light and water availability.

Materials and methods

Plant material and microclimate growing conditions

Seedlings of *N. pumilio*, 2-3 years old, measuring 6-7 cm in height, were obtained from the understory of mature forests (54° 06' S, 68° 37' W). Seedlings were collected in September, at the beginning of the spring season in the Southern Hemisphere, before budburst, from stands with high canopy cover (94 ± 5%). Seedlings were transplanted into plastic pots (14 cm diameter and 15 cm height) filled with a substrate of peat/sand/forest soil (1:1:1). The substrate corresponded to a clay loam soil (sand-silt-clay, 36-24-40%) with 7% organic matter, pH 4.99, and soil field capacity of 81% (Martínez Pastur et al. 2007).

Incident solar radiation during the growing season (October to March) averaged $2085 \pm 534 \mu\text{mol m}^{-2} \text{s}^{-1}$, with a maximum absolute value of $2702 \mu\text{mol m}^{-2} \text{s}^{-1}$. Plants were grown in a greenhouse in Ushuaia city (Tierra del Fuego – 54° 46' S, 68° 12' W) during one growing season, under a 100

(Peri et al. 2009).

Following tree logging, seedling survival and rapid growth depend on their ability to adjust to new microclimatic conditions (Tognetti et al. 1998, Toro-Manríquez et al. 2019, Mestre et al. 2024).

While previous studies have characterized the growth and gas exchange patterns of *N. pumilio* under different regimes (Lencinas et al. 2007, Martínez Pastur et al. 2011b), the underlying photochemical mechanism remains unexplored. Understanding the interaction between light and water availability helps explain how soil moisture stress significantly modulates a plant's ability to process light energy (Hikosaka 2021). When soil moisture is excessive, metabolic limitations and reduced nutrient uptake often decrease CO_2 assimilation rates, creating a "sink limitation". In these conditions, absorbed light energy exceeds the processing capacity of the pho-

tosynthetic apparatus, potentially intensifying photo-oxidative stress under the high irradiance levels typical of canopy gaps (Murchie & Lawson 2013).

To evaluate these complex responses, we combined chlorophyll fluorescence and pigment analysis. Both approaches provide complementary information about plants' eco-physiological adaptation capacity. Chlorophyll fluorescence reveals the real-time operational efficiency of the photosynthetic apparatus, whereas pigment content reflects dynamic acclimatory responses over short- to intermediate-timescales. Specifically, adjustments in light-harvesting pigments (chlorophyll *a* and *b*) and photoprotective pigments (carotenoids) involve structural and biochemical regulation in response to specific environmental conditions. Assessing these factors is essential to distinguish between short-term functional resilience and structural

μm commercial nylon cover to prevent natural rainfall. Three light intensity treatments were established using zero, one, or two layers of commercial black shade cloth: high ($1334.4 \pm 235.82 \mu\text{mol m}^{-2} \text{s}^{-1}$, i.e., 64% of natural irradiance), medium ($542.1 \pm 74.65 \mu\text{mol m}^{-2} \text{s}^{-1}$, 26%), and low ($83.4 \pm 61.97 \mu\text{mol m}^{-2} \text{s}^{-1}$, 4%). The low-light-intensity treatment simulated the light conditions typically found in primary forests, while the medium- and high-intensity treatments represented conditions typical of harvested stands with varying canopy opening (see Martínez Pastur et al. 2011b for more details).

Temperature was controlled via forced ventilation to ensure that canopy-level temperature did not exceed 24°C . Air humidity, as well as air and soil temperatures, was continuously monitored in each light-intensity treatment using data loggers throughout the study period (Fig. S1 in Supplementary material). Soil water content was actively managed through manual irrigation to maintain two distinct treatment levels: half of the plants were kept at 40–60% of soil field capacity (optimal soil moisture), while the other half were maintained at 80–100% (excess soil moisture), representing a potential stress condition. For each combination of light level and soil moisture, six blocks of 40 plastic pots were allocated in distinct positions within the greenhouse (Fig. 1). Irrigation in the greenhouse varied according to the combination of light and soil moisture treatments (six treatments in total) and was controlled gravimetrically every 3 days throughout the growing season in response to seasonal changes in temperature and evaporative demand. Pot weight was monitored, and when pots approached the lower threshold of each soil moisture treatment (40% for optimal moisture and 80% for excess moisture), they were irrigated up to the upper limit of the corresponding range. Treatments under low and medium light levels required lower irrigation frequency, with 13 to 23 irrigation events applied throughout the growing season, whereas treatments under high light intensity required more frequent irrigation to keep pots within the target soil moisture range (36 to 45 irrigation events). Most of the seedlings survived transplanting, with a mortality rate of 0.5% to 2.4% in spring. During the first week of each month of the growing season (October to March), six plastic pots per block per treatment were randomly chosen for measurements (6 seedlings $\times$ 6 treatments $\times$ 6 months, $n = 216$).

Sampling and measurements

Phenological status was monitored monthly from the beginning of spring (September) to the beginning of autumn (March). Each month, 30 plants were randomly selected from each treatment, and their phenological stages were recorded, including budburst, leaf unfolding, full leaf

expansion, appearance of first red leaves, red leaves, and complete leaf fall. We also recorded the number of leaves per plant as a proxy for vegetative growth. This metric has been used as an indicator of seedling vigor and meristematic activity in response to environmental stress (Pérez-Harguindeguy et al. 2013).

For chlorophyll fluorescence measurements, randomly selected pots were moved to controlled laboratory conditions ($12\text{--}14^\circ\text{C}$ in complete darkness) for an 8-hour acclimation period before measurements. Chlorophyll fluorescence was assessed in the first fully expanded, intact leaf that remained attached to the stem. Measurements were taken on one leaf per plant, with six plants per treatment ($n = 6$ leaves per treatment), using a Chlorophyll fluorometer (Model S151, Qubit Systems, Kingston, Ontario, Canada). This instrument uses a pulsed LED light source (peak wavelength 660 nm, 50 Hz) to excite chlorophyll fluorescence without inducing photosynthesis. Emitted fluorescence is detected through a long-pass filter ($>700 \text{ nm}$), ensuring that only the fluorescence emitted by the leaf in response to the pulsed LED is measured. This setup enables precise detection across a wide range of irradiance levels, from complete darkness to saturated light conditions. Each leaf was exposed to a saturation pulse of high light intensity ($2250 \mu\text{mol m}^{-2} \text{s}^{-1}$) for five seconds to determine the following fluorescence variables: fluorescence in the absence of photosynthetic light (F_0), which occurs when all PSII reaction centres are open; maximum fluorescence level (F_m), which occurs when all reaction centres are closed; variable fluorescence (F_v), that is the difference between F_0 and F_m ; maximum fluorescence under illuminated conditions (F'_m); steady-state yield of fluorescence in the light (F_s); and zero level fluorescence in the light (F'_0). From chlorophyll fluorescence parameters, the following coefficients were calculated: the effective quantum yield of PSII photochemistry ($\Phi_{\text{PSII}} = [F'_m - F_t] / F'_m$ – Maxwell & Johnson 2000); photochemical quenching ($qP = [F'_m - F_t] / [F'_m - F_0]$ – Genty et al. 1989); maximum quantum yield of PSII in dark-adapted leaves (F_v/F_m – Butler 1978); and non-photochemical quenching ($\text{NPQ} = [F_m/F'_m] - 1$ – Bilger & Björkman 1990). These standard parameters are widely used to assess photosynthetic responses, such as diurnal photoinhibition and water stress (Chen et al. 2022), and to model light-response curves of Φ_{PSII} (Yang et al. 2025).

Complementarily, chlorophyll and carotenoid contents were determined by a spectrophotometric method applied in Lichtenhaler & Babani (2022). Fresh leaf discs (0.5 cm^2) were obtained from the first fully expanded leaf of the same plants where chlorophyll fluorescence was measured, and processed immediately after collection. Each leaf disc was placed in 2 ml of an aqueous acetone-Tris buffer solution (80%

v/v, pH 7.8). This solution was centrifuged at $1610 \times g$ for 10 minutes at 20°C . The supernatant was used to determine absorbance at 470, 537, 647 and 663 nm with a spectrophotometer to quantify pigment concentrations and contents according to Sims & Gamon (2002), including chlorophyll *a* (ChlA, mmol m^{-2}), chlorophyll *b* (ChlB, mmol m^{-2}), total chlorophyll (ChlT, mmol m^{-2}), ratio between chlorophyll *a* and *b* (ChlA/ChlB), carotenoids (Car, mmol m^{-2}), and ratio between total chlorophyll and carotenoids (ChlT/Car).

Statistical analyses

For each response variable (number of leaves, fluorescence and pigment variables), a three-way analysis of variance (ANOVA) was performed, with month (October to March), light intensity (4%, 26%, and 64% of the natural incident irradiance) and soil moisture (40–60%, and 80–100% soil field capacity) as fixed factors, including all possible interactions among factors. Because most variables showed significant interactions, we analyzed simple effects by performing post hoc comparisons for each factor at each level of the interacting factor (e.g., light intensity within each month) using Tukey's multiple range test ($\alpha = 0.05$). Statistical analyses were performed in R (R Core Team 2018).

Correlation analyses were conducted among the response variables to evaluate their relationships. Principal Component Analyses (PCA) were performed on a matrix of 36 rows and 7 columns to rank and visualize multivariate responses of plants by integrating pigment-related variables with chlorophyll fluorescence parameters across treatments at three representative periods of the growing season: early spring (October), mid-summer (January), and late summer/early autumn (March). A subset of eco-physiological variables was selected for the PCA analyses to reduce redundancy among correlated traits, prioritizing variables with stronger contributions to the principal components and clearer treatment-related responses. Correlation analyses and PCAs were performed using the “vegan” package (Oksanen et al. 2025) in R (R Core Team 2018).

Results

Phenology and leaf production

The phenological status of the seedlings defined the study period, which extended from October to March (Fig. 2). At the beginning of spring (September), all seedlings were at the budburst stage, followed by leaf unfolding during October and November. From December onwards, distinct patterns emerged in both the rate of leaf development and the total number of leaves. Seedlings reached an early plateau in leaf number by early December across three treatments (the two low-light conditions and the high-light-excess-soil-moisture treatment). Consequently, in these

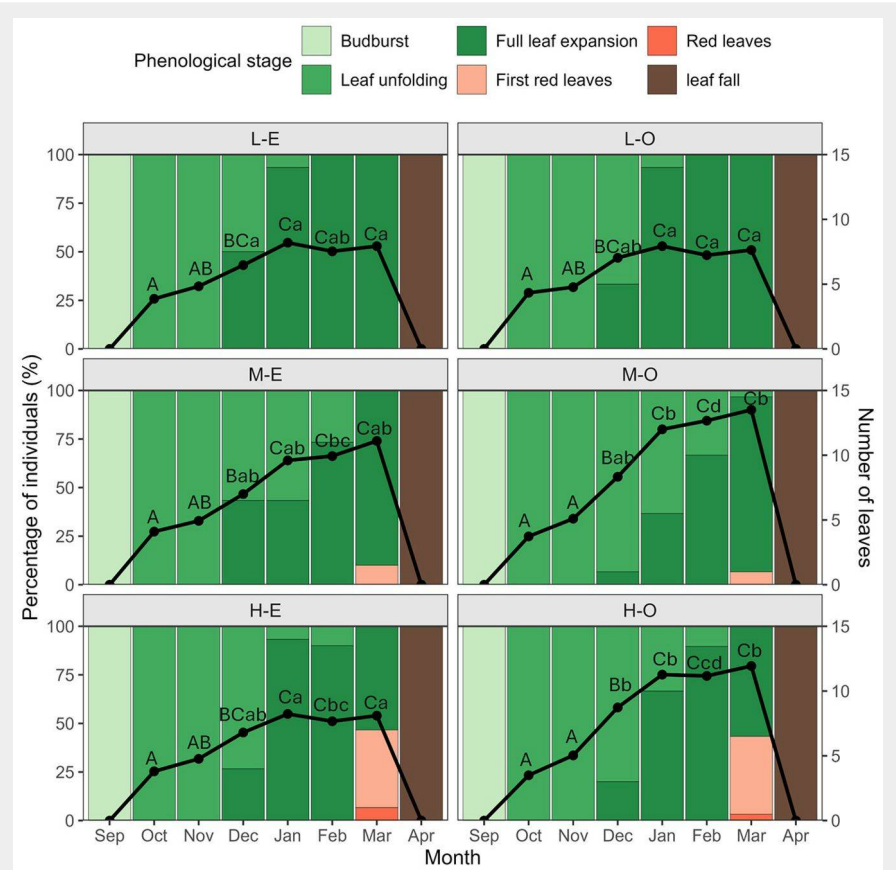


Fig. 2 - Seasonal progression of phenological stages (stacked bars, left Y-Axis) and leaf number (black line and points, right Y-Axis) in *Nothofagus pumilio* seedlings under the six studied treatments (combinations of light intensity and soil moisture), representing the interactions shown in Tab. 1. Different capital letters indicate significant differences among months within treatments, whereas lowercase letters indicate significant differences among treatments within each month (Tukey test, $p < 0.05$). (L-E): low light intensity $\times$ excess soil moisture; (L-O): low light intensity $\times$ optimal soil moisture; (M-E): medium light intensity $\times$ excess soil moisture; (M-O): medium light intensity $\times$ optimal soil moisture; (H-E): high light intensity $\times$ excess soil moisture; (H-O): high light intensity $\times$ optimal soil moisture.

treatments, the highest proportion of leaves was fully expanded in January, and leaf number remained stable up to the end of the growing season (Tab. 1, Fig. 2). In contrast, leaf number continued to increase significantly after December in the other treatments, resulting in a higher proportion of plants still bearing expanding

leaves during this period. Red leaves appeared early under medium and high light intensities (March), with the highest proportion in the high-light treatments. The final leaf number was significantly higher under medium and high light with optimal soil moisture (Tab. 1, Fig. 2). By April, all treatments showed complete leaf loss.

Tab. 1 - ANOVA results for the number of leaves in *Nothofagus pumilio* seedlings along the growing season, considering month, light intensity, and soil moisture levels as factors and their interactions. (df): Degrees of freedom.

Effects	df	Number of leaves	
		F statistics	p-value
Month	5	126.48	<0.001
Light	2	38.21	<0.001
Moisture	1	38.28	<0.001
Month: Light	10	6.33	<0.001
Month: Moisture	5	3.97	0.002
Light: Moisture	2	10.14	<0.001
Month: Light: Moisture	10	1.78	0.067

Chlorophyll fluorescence

All raw fluorescence parameters (F_0 , F_m , F'_m , F_v , and F'_v) were significantly affected by the three main factors studied, with multiple significant interactions (Tab. S1 in Supplementary material), indicating a complex response at the signal level. In contrast, the derived physiological coefficients (Φ_{PSII} , qP , F_v/F_m , and NPQ) were driven by simpler dynamics, primarily affected by seasonal variation and light intensity, whereas soil moisture had no significant effect on the main physiological coefficients (Tab. 2). Light intensity modulated the temporal dynamics of three coefficients (Φ_{PSII} , qP , and NPQ), generating distinct seasonal patterns (Tab. 2, Fig. 3). F_v/F_m was the most stable parameter to the influence of light and soil moisture, varying only across the season ($p < 0.01$).

The efficiency of PSII photochemistry (Φ_{PSII}) and photochemical quenching (qP) showed a consistent seasonal pattern, with low values at the beginning of the growing season, a peak during mid-season, and a decline towards the end of the growing season. This reflects the seasonal trajectory of photosynthetic capacity, from initial development through peak performance to senescence. Light intensity modulated the temporal trend, reducing photochemical efficiency in late summer via photoinhibition or greater reliance on photoprotective mechanisms under high irradiance (Fig. 3). Non-photochemical quenching (NPQ) showed contrasting responses depending on light conditions, reflecting shifts in photoprotective strategies. Under low light, NPQ was higher at the beginning of the season, whereas under high light, it increased during mid- to late season, indicating enhanced energy dissipation at higher irradiance (Fig. 3).

Photosynthetic pigments content

Photosynthetic pigment content in *Nothofagus pumilio* seedlings exhibited clear acclimation to light levels throughout the growing season, while also being influenced by soil moisture (Tab. 3). Chlorophyll *a* (Chl*A*), chlorophyll *b* (Chl*B*), and total chlorophyll (Chl*T*) were strongly and positively correlated with each other (Tab. S3 in Supplementary material) and varied according to the interactions between month $\times$ light intensity and between month $\times$ soil moisture. Treatment effects on pigment concentrations were not evident at the onset of the growing season. However, as the season progressed, clear differences emerged across light treatments. Seedlings under low light initially accumulated higher chlorophyll concentrations, whereas from mid-season onwards, those under medium light consistently exhibited the highest values, reaching peak levels towards the end of the growing season (Fig. 4). Differences in pigment composition were also observed, with higher Chl*B* under low light compared to high light, while Chl*A* showed less consistent variation among treatments

Effects	df	ΦPSII		qP		F _v /F _m		NPQ	
		F-stat	p-value	F-stat	p-value	F-stat	p-value	F-stat	p-value
Month	5	56.6	<0.001	50.1	<0.001	5.8	<0.001	18.9	<0.001
Light	2	1.7	0.193	0.9	0.415	0.7	0.488	5.9	0.003
Moisture	1	2.9	0.091	3.8	0.052	0.2	0.628	0.2	0.628
Month × Light	10	2.1	0.029	2.4	0.01	1.0	0.413	3.9	<0.001
Month × Moisture	5	0.6	0.722	0.8	0.553	0.7	0.592	1.8	0.119
Light × Moisture	2	0.9	0.394	0.5	0.615	1.6	0.205	0.2	0.795
Month × Light × Moisture	10	0.6	0.831	0.5	0.871	1.1	0.357	1.4	0.178

was less consistent, indicating a weaker differentiation among light treatments. The total chlorophyll-to-carotenoid ratio (ChlT/Car) showed the opposite trend, with higher values at low and medium light intensities than at high light across both soil moisture conditions. This pattern reflects the combined effect of reduced carotenoid content under low light and increased chlorophyll content under intermediate light (Fig. 4, Fig. 5). Together, these patterns indicate coordinated adjustments in pigment composition, with increasing light intensity

Light intensity: High (red circle), Medium (blue circle), Low (black circle)

PhiPSII

A	AB	BCD	D	CD	BC
A	B	C	C	C	BC
A	AB	BC	C	BC	BC

qp

A	AB	BCD	D	CD	BC
A	B	C	C	C	BC
A	AB	BC	D	D	CD

F_v/F_m

A	B	B	B	B	B
B	B	A	A	A	A
A	B	AB	AB	AB	AB

NPQ

B	A	A	A	A	A
B	B	A	A	A	A
A	B	AB	AB	AB	AB

Tab. 3 - ANOVA results of pigment content variables in *Nothofagus pumilio* seedlings along the growing season, considering month, light intensity, and soil moisture levels as factors and their interactions. (ChIA): chlorophyll *a* (mmol m⁻²); (ChIB): chlorophyll *b* (mmol m⁻²); (ChIT): total chlorophyll (mmol m⁻²); (ChIA/ChIB): ratio between chlorophyll *a* and *b*; (Car): carotenoids (mmol m⁻²); (ChIT/Car): ratio between total chlorophyll and carotenoids; (df): degrees of freedom; (F-stat): F statistics.

Effect	df	ChIA		ChIB		ChIT		Car		ChIA/ChIB		ChIT/Car	
		F-stat	p-value	F-stat	p-value	F-stat	p-value	F-stat	p-value	F-stat	p-value	F-stat	p-value
Month	5	81.3	<0.001	65.2	<0.001	75.1	<0.001	34.8	<0.001	83.8	<0.001	14.2	<0.001
Light	2	14.8	0.000	58.7	<0.001	20.0	<0.001	261.4	<0.001	18.6	<0.001	94.7	<0.001
Moisture	1	4.3	0.040	3.8	0.052	2.7	0.101	0.1	0.760	0.4	0.513	5.8	0.017
Month: Light	10	5.9	<0.001	7.3	<0.001	4.7	<0.001	52.0	<0.001	4.0	<0.001	5.0	<0.001
Month: Moisture	5	2.7	0.022	3.1	0.010	3.8	0.002	1.3	0.250	3.1	0.010	3.2	0.008
Light: Moisture	2	1.1	0.347	0.5	0.631	0.4	0.685	2.6	0.08	1.4	0.254	0.9	0.428
Month: Light: Moisture	10	1.3	0.210	1.4	0.194	1.5	0.143	2.0	0.040	2.4	0.010	3.5	<0.001

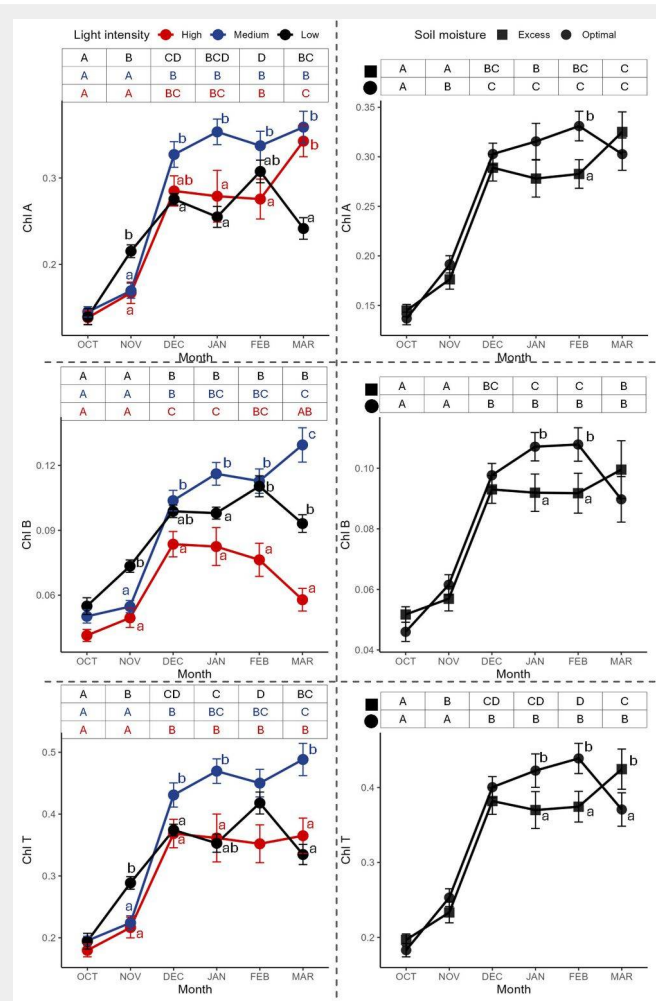


Fig. 4 - Leaf tissue pigment content of chlorophyll *a* (ChIA), chlorophyll *b* (ChIB), and total chlorophyll (ChIT) in *Nothofagus pumilio* seedlings over the period studied and in three light intensity and two soil moisture treatments, representing the interactions shown in Tab. 3. When light intensity × month is shown, the data are averages of the two soil moisture treatments. When soil moisture × month is shown, the data are averages of the three light treatments. Error bars represent the standard error. Values followed by different capital letters represent significant differences ($p < 0.05$) among months for each light intensity (or soil moisture) treatment, while lower-case letters represent significant differences among light intensity (or soil moisture) levels for each month, according to the Tukey test.

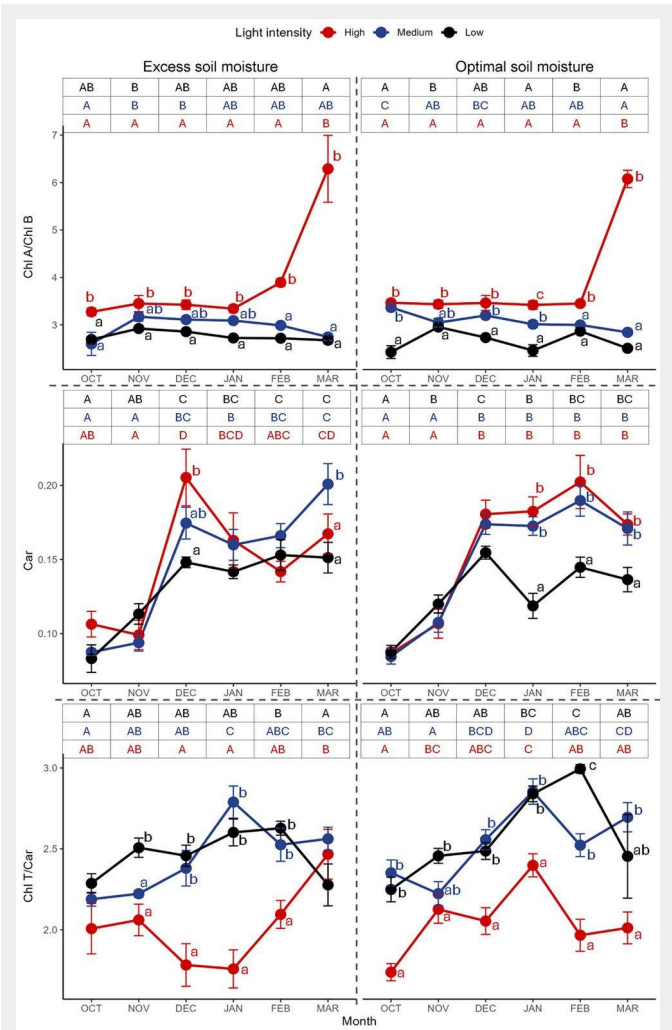


Fig. 5 - Ratio between chlorophyll *a* and *b* (ChIA/ChIB), carotenoids (Car, mmol m⁻²), and total chlorophyll-to-carotenoid ratio (ChIT/Car) in *Nothofagus pumilio* seedlings over the study period under three light intensity treatments and two soil moisture levels. Panels represent the triple interactions shown in Tab. 3, separated by soil moisture treatment. Values followed by different capital letters represent significant differences ($p < 0.05$) among months for each light intensity and soil moisture treatment, while lower-case letters represent significant differences ($p < 0.05$) among light intensity levels for each month and soil moisture treatment, according to the Tukey test.

promoting a shift from light-harvesting investment (higher chlorophyll content and lower ChlA/ChlB ratio) towards enhanced photoprotection (higher carotenoid content and lower ChlT/Car ratio). In turn, soil moisture modulated this response, with optimal soil moisture enhancing pigment accumulation and photoprotective adjustments.

Covariation between chlorophyll fluorescence and photosynthetic pigments

Chlorophyll fluorescence-based photosynthetic performance was moderately associated with pigment content (Fig. S2 in Supplementary material). NPQ was negatively correlated with pigment content, whereas Φ PSII and qP showed positive relationships, particularly with total chlorophyll content (ChlT). This relationship pattern reflects the multivariate association patterns of seedlings across different treatments and key moments of the growing season (Fig. 6). PCA ordinations integrated physiological and biochemical traits into coordinated patterns of trait covariation, revealing a consistent multivariate structure of the photosynthetic apparatus. PC1 (37–42% of explained variance) was predominantly associated with chlorophyll content, with ChlA and ChlB contributing most (Tab. S2). PC2 (26–31%) captured a secondary gradient of variation related to pigment ratios (ChlA/ChlB and ChlT/Car), carotenoid content, and NPQ, and showed greater temporal variability. Across seasons, treatment separation was primarily driven by light availability: high-light conditions were clearly separated across the three periods, whereas medium and low light were also distinguishable but showed greater overlap at the beginning of the season. In contrast, soil moisture had a weaker effect, with differences occurring mainly within each light treatment rather than generating a distinct overall separation pattern. This pattern was most pronounced in mid-summer, when treatment groups were clearly differentiated. By March, treatment effects were still apparent but less distinct, with increased overlap among groups, particularly between soil moisture levels within the same light treatments. This suggests a seasonal convergence in seedling physiological responses towards the end of the growing season, while the main gradients associated with light and pigment composition remain detectable.

Discussion

Seasonal phenology and adjustment of the photosynthetic machinery

The leaf phenology of *N. pumilio* seedlings under greenhouse conditions was largely consistent with the natural growth patterns described for the species, although leaf formation in natural forests typically peaks in early summer (Premoli et al. 2007, Mondino et al. 2019). The early

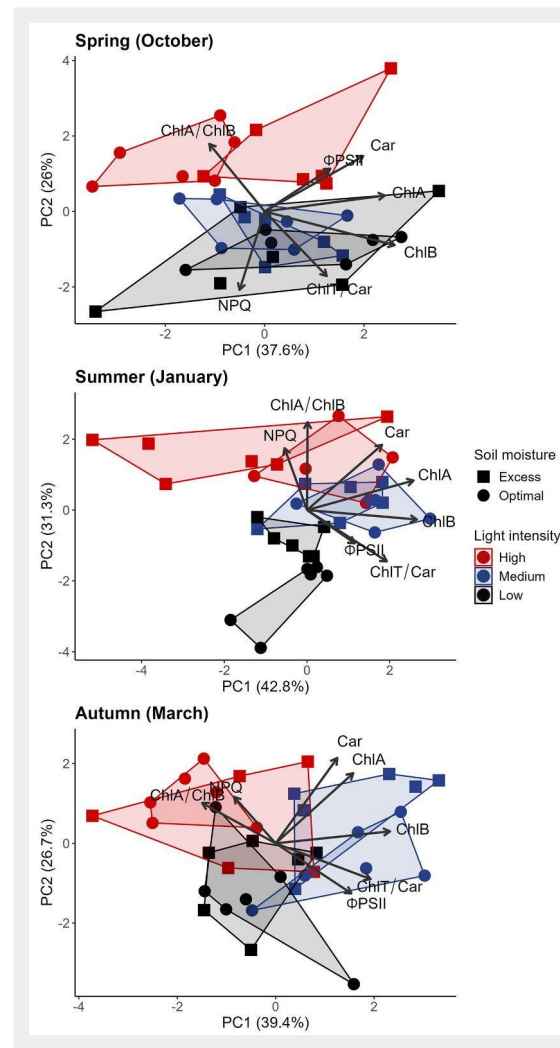


Fig. 6 - Principal component analyses (PCA) integrating chlorophyll fluorescence parameters and photosynthetic pigment contents of *Nothofagus pumilio* seedlings at three representative periods of the growing season: early spring (October), mid-summer (January), and early autumn (March). Each PCA represents the multivariate response of seedlings under the different combinations of light intensity and soil moisture treatments. Points correspond to individual plants.

plateau in leaf number observed under excess soil moisture and high irradiance reflects a developmental constraint, likely due to environmental stress (either deep shade or soil saturation) that accelerates the leaf phenological cycle at the expense of total structural growth, mirroring the reduced biomass reported by Lencinas et al. (2007).

The transition from the shaded understory to the high-irradiance environment of canopy gaps represents a critical bottleneck for *Nothofagus pumilio* regeneration (Toro-Manríquez et al. 2019). Our results reveal that seedlings navigate this transition through a differential allocation strategy, prioritizing the maintenance of photosynthetic function over structural investment. While light intensity dictated the metabolic “pathway” (photoprotection vs. light-harvesting), high soil moisture significantly constrained the magnitude of this investment, as shown by the observed interactions (Fig. 3, Fig. 4, Fig. 5). At the same time, PCA ordinations (Fig. 6) reinforced the central role of light availability as the main driver of seedling eco-physiological differentiation throughout the growing season. This is consistent with our hypothesis that physiological performance in *N. pumilio* seedlings follows a light-dependent

response modulated by soil moisture availability. While a generalized synergistic effect was expected, the analysis indicates that this interaction is trait-dependent: synergistic negative effects occur between high light and excess soil moisture, primarily expressed in structural traits, whereas functional responses remain comparatively stable. This partial decoupling allowed seedlings to remain physiologically operational even when structural development (leaf number and pigment synthesis) was impaired by excess soil moisture or low light intensity, highlighting the prioritization of functional integrity under stress conditions.

PSII efficiency and energy dissipation mechanisms (quenching)

The actual efficiency of PSII photochemistry (Φ PSII) is modulated by the balance between energy capture and metabolic utilization (Murchie & Lawson 2013). The results show that Φ PSII remained stable across treatments until late in the growing season, with declines under high light conditions, consistent with previous evidence that, under stress, Φ PSII decreases more rapidly as light intensity increases (Zait et al. 2024). A similar response for photochemical quenching (qP) reflects the pro-

portion of open PSII reaction centers. The reduction in qP under medium and high light conditions indicates limited use of absorbed energy, likely associated with light saturation (Murchie & Ruban 2020).

The sustained high F_v/F_m values (about 0.80) indicate no irreversible damage to PSII (Mayoral et al. 2015, Krzeminska et al. 2024). The significant decrease in both Φ_{PSII} and qP in the high-light treatment indicates photochemical downregulation, a state known as dynamic photoinhibition (Murchie & Ruban 2020). The stability of F_v/F_m therefore demonstrates the efficacy of photoprotective mechanisms, primarily high NPQ, in safely dissipating this excess energy. This prevented acute downregulation from escalating into chronic, damaging photoinhibition (Takahashi & Murata 2008). Our results for *N. pumilio* were similar to those reported for *Fagus* by Cascio et al. (2010) and Desotgiu et al. (2012), which also showed no significant differences in F_v/F_m across different light conditions.

High NPQ under elevated irradiance indicates a shift toward dissipating thermal energy when light supply exceeds photosynthetic capacity, serving as a key photoprotective mechanism against excess light (Mattila & Tyystjärvi 2023). The higher NPQ activity in expanding leaves at the beginning of the season could be related to leaf youth (ontogeny), as developing chloroplasts and lower photosynthetic capacity require greater thermal dissipation to avoid photo-oxidative stress (Hieke et al. 2002, Bielczynski et al. 2017). While we detected a positive correlation between photochemical efficiency (Φ_{PSII} , qP , F_v/F_m) and pigment content (Fig. S2 in Supplementary material), it was not as strong as correlations reported in other studies (Torres Netto et al. 2005). Our results for photochemical parameters did not support the hypothesis of a synergistic interaction between stressors, as photochemical performance was primarily driven by light intensity rather than soil moisture, allowing seedlings to maintain functional stability across the two soil moisture levels analyzed.

Photosynthetic pigment content adjustments

Consistent with the literature, pigment content acclimation to light levels in *N. pumilio* followed predictable patterns: photodegradation explains the reduction in total chlorophyll under high irradiance (Tian et al. 2021), whereas the lower chlorophyll a/b ratio under shade represents an optimization of light harvesting (Taiz et al. 2023). Thus, intermediate light conditions represent an optimal balance for pigment accumulation in this species, avoiding both light limitation under shade and photodegradation under high irradiance.

Reduced pigment concentrations under saturated soil conditions during peak growth reflect root-level stress, given that excess soil moisture has been shown to im-

pair root development in *N. pumilio* (Lencinas et al. 2007, Martínez Pastur et al. 2011b). Such soil saturation hinders nutrient uptake and, consequently, pigment synthesis (Voeselek & Bailey-Serres 2015), particularly in species such as *N. pumilio*, which is adapted to well-drained, poorly developed soils (Van Galen et al. 2021, Mestre et al. 2024). Moreover, Peri et al. (2009) reported that *N. pumilio* rapidly reduces its normalized net photosynthetic rate under waterlogging conditions, further supporting the species' sensitivity to soil saturation. Furthermore, high irradiance intensifies pigment destruction because the plant cannot synthesize new photosynthetic and photoprotective compounds (e.g., carotenoids) quickly enough (Döweler et al. 2021). Carotenoids play a dual role, acting as accessory pigments in light harvesting and as protectors against photooxidative damage, protecting chlorophylls from destructive reactions (Lichtenhaler 1987, Lichtenthaler et al. 2007, Rojas-Rioseco et al. 2023).

Our results reveal a clear distinction between the temporal dynamics of functional and structural traits. Photosynthetic function followed a seasonal divergence-convergence pattern, peaking in January and February before converging in March. This discrepancy reinforces the decoupling between function and structure: while photochemical efficiency is an immediate response that declines during senescence, pigment profiles represent a more permanent strategic investment. The persistence of treatment-specific pigment ratios ($ChlA/ChlB$) towards the end of the growing season indicates that structural adjustments outlast photochemical responses, reflecting a lasting imprint of the environmental conditions experienced by seedlings. This appears to reflect a strategic acclimation, a structural adjustment that integrates resource availability with the plant's broader phenological context.

Phenotypic plasticity: management implications and research perspectives

The mechanisms described above exemplify the high phenotypic plasticity of *N. pumilio*, which may contribute to its ability to cope with disturbed environments. Overall, this study demonstrates a clear acclimation of *N. pumilio* seedlings to light treatments influenced by soil moisture throughout the growing season. Plants preconditioned under a closed canopy rapidly adjusted their photosynthetic physiology (chlorophyll fluorescence and pigment content) in response to the new light regimes and soil moisture conditions. This acclimatization capacity is essential for regeneration survival after canopy opening by silvicultural cutting. Our study suggests that while seedlings can tolerate high irradiance through efficient photoprotective mechanisms (as evidenced by high NPQ and carotenoid accumulation), medium-light conditions appear most favorable for

plants during their first years of life. Under moderate light levels, seedlings maximized chlorophyll content while maintaining PSII integrity. These findings align with previous reports of *N. pumilio* seedling plasticity in physiological variables such as photosynthetic rates (Martínez Pastur et al. 2007) and morphological traits (Lencinas et al. 2007, Martínez Pastur et al. 2011b). However, these findings should be contextualized. While this study evaluated the plant response over a single growing season, plasticity is a gradual process that develops over multiple seasons (Escobar-Sandoval et al. 2021), and its adaptive advantages are also shaped by ecological context and competition with other colonizing species. Furthermore, the optimal soil moisture conditions in this study (well-drained, 50-60% field capacity) reflect the high-quality sites of primary forests (Martínez Pastur et al. 2007). In contrast, the excess moisture treatment reflects the post-harvesting edaphic conditions in low-lying areas near wet meadows and peatlands, where drainage is naturally restricted (Grootjans et al. 2010), or the novelty created by the expansion of the invasive beaver (*Castor canadensis* – Henn et al. 2016). Therefore, to gain a comprehensive understanding, it is necessary to contrast these results with other potential environmental scenarios, such as rising temperatures and reduced soil moisture, across a comparable canopy-cover gradient to that analyzed in this study. Evaluating seedling responses under different scenarios, particularly in the context of climate change, would help determine whether the observed plasticity is sufficient to buffer the effects of a more extreme climate, a critical consideration for the long-term conservation and management of these forests.

Conclusions

Nothofagus pumilio seedlings show high phenotypic plasticity, enabling to adjust their photosynthetic apparatus during a single growing season in response to changes in light and soil moisture, conditions that often occur after timber harvesting. Seedling development early in the season is optimal under intermediate light conditions. Although seedlings can tolerate high irradiance through photoprotective mechanisms, partial canopy cover allows them to maximize photosynthetic performance without suffering chronic stress. This acclimation strategy could be supported by a differential physiological response, in which seedlings rely on rapid, short-term functional regulation (photochemical efficiency) to cope with light fluctuations, while their structural adjustment investment (pigment synthesis) reflects a strategic adjustment that integrates more complex, chronic environmental stress factors, including soil moisture. This physiological explanation supports the recommendation to adopt silvicultural practices that maintain partial canopy cover to pro-

mote successful regeneration. By promoting stable, intermediate light conditions, the establishment and survival of seedlings with a resilient physiological state is encouraged, which is key to ensuring the persistence of forests in the face of increasing climate variability.

List of abbreviations

The following abbreviations have been used throughout the paper:

- F_0 : Fluorescence in the absence of photosynthetic light;
- F_m : Maximum fluorescence level;
- F_v : variable fluorescence;
- F'_m : Maximum fluorescence in the light;
- F_s : Steady-state yield of fluorescence in the light;
- F'_0 : Zero-level fluorescence in the light;
- $\Phi PSII$: Efficiency of PSII photochemistry;
- qP : Photochemical quenching;
- F_v/F_m : Maximum quantum yield of PSII;
- NPQ: Non-photochemical quenching;
- ChlA: Chlorophyll a;
- ChlB: Chlorophyll b;
- ChlT: Total chlorophyll;
- ChlA/ChlB: Ratio between chlorophyll a and b;
- Car: Carotenoids;
- ChlT/Car: Ratio between total chlorophyll and carotenoids.

Acknowledgments

The authors gratefully acknowledge Emilce Gallo and Enrique Barrio for their collaboration during data collection and sampling processing.

Funding Declaration

This work was partly supported by PAE2004 22428 (ANPCyT, Argentina) and BOSAMCA MIA (CATIE, Costa Rica).

References

- Bielczynski LW, Lacki MK, Hoefnagels I, Gambin A, Croce R (2017). Leaf and plant age affect photosynthetic performance and photoprotective capacity. *Plant Physiology* 175: 1634-1648. - doi: [10.1104/pp.17.00904](https://doi.org/10.1104/pp.17.00904)
- Bilger W, Björkman O (1990). Role of the xanthophyll cycle in photoprotection elucidated by measurements of light-induced absorbance changes, fluorescence and photosynthesis in leaves of *Hedera canariensis*. *Photosynthesis Research* 25 (3): 173-185. - doi: [10.1007/BF00033159](https://doi.org/10.1007/BF00033159)
- Butler WL (1978). Energy distribution in photochemical apparatus of photosynthesis. *Annual Review of Plant Physiology* 29: 345-378. - doi: [10.1146/annurev.pp.29.060178.002021](https://doi.org/10.1146/annurev.pp.29.060178.002021)
- Cascio C, Schaub M, Novak K, Desotgiu R, Bussotti F, Strasser RJ (2010). Foliar responses to ozone of *Fagus sylvatica* L. seedlings grown in shaded and in full sunlight conditions. *Environmental and Experimental Botany* 68: 188-197. - doi: [10.1016/j.envexpbot.2009.10.003](https://doi.org/10.1016/j.envexpbot.2009.10.003)
- Chaves JE, Rodríguez-Souilla J, Cellini JM, Lencinas MV, Peri PL, Martínez Pastur G (2024). Shelterwood cut intensity determines recovery pathways of managed *Nothofagus pumilio* forests. *New Zealand Journal of Forestry Science* 54. - doi: [10.33494/nzjfs542024x301x](https://doi.org/10.33494/nzjfs542024x301x)
- Chen Z, Liu Z, Han S, Jiang H, Xu S, Zhao H, Ren S (2022). Using the diurnal variation characteristics of effective quantum yield of PSII photochemistry for drought stress detection in maize. *Ecological Indicators* 138: e108842. - doi: [10.1016/j.ecolind.2022.108842](https://doi.org/10.1016/j.ecolind.2022.108842)
- Cuevas J, Arroyo MK (1999). Ausencia de banco de semillas persistente en *Nothofagus pumilio* (Fagaceae) en Tierra del Fuego, Chile [Absence of a persistent seed bank in *Nothofagus pumilio* (Fagaceae) in Tierra del Fuego, Chile]. *Revista Chilena de Historia Natural* 72: 73-82. [in Spanish]
- Desotgiu R, Pollastrini M, Cascio C, Gerosa G, Marzuoli R, Bussotti F (2012). Chlorophyll a fluorescence analysis along a vertical gradient of the crown in a poplar (Oxford clone) subjected to ozone and water stress. *Tree Physiology* 32: 976-986. - doi: [10.1093/treephys/tps062](https://doi.org/10.1093/treephys/tps062)
- Donoso PJ, Promis A, Loguerio GA, Attis Beltrán H, Caselli M, Chauchard LM, Cruz G, González Peñalba M, Martínez Pastur G, Navarro C, Núñez P, Salas-Eljatib C, Soto DP, Vásquez-Grandón A (2022). Silviculture of South American temperate native forests. *New Zealand Journal of Forestry Science* 52: 1-32. - doi: [10.33494/nzjfs522020x173x](https://doi.org/10.33494/nzjfs522020x173x)
- Döweler F, Case BS, Buckley HL, Bader MKF (2021). High light-induced photoinhibition is not limiting seedling establishment at abrupt tree-line ecotones in New Zealand. *Tree Physiology* 41 (11): 2034-2045. - doi: [10.1093/treephys/tpa4061](https://doi.org/10.1093/treephys/tpa4061)
- Escobar-Sandoval M, Pâques L, Fonti P, Martínez-Meier A, Rozenberg P (2021). Phenotypic plasticity of European larch radial growth and wood density along a 1,000 m elevational gradient. *Plant-Environment Interactions* 2: 45-60. - doi: [10.1002/pei3.10040](https://doi.org/10.1002/pei3.10040)
- Genty B, Briantais JM, Baker NR (1989). The relationship between the quantum yield of photosynthetic electron transport and quenching of chlorophyll fluorescence. *Biochimica et Biophysica Acta - General Subjects* 990: 87-92. - doi: [10.1016/S0304-4165\(89\)80016-9](https://doi.org/10.1016/S0304-4165(89)80016-9)
- Gustafsson L, Baker SC, Bauhus J, Beese WJ, Brodie A, Kouki J, Lindemayer DB, Lohmus A, Martínez Pastur GJ, Messier C, Neyland M, Palik B, Sverdrup-Thygeson A, Volney WJA, Wayne A, Franklin JF (2012). Retention forestry to maintain multifunctional forests: a world perspective. *BioScience* 62: 633-645. - doi: [10.1525/bio.2012.62.7.6](https://doi.org/10.1525/bio.2012.62.7.6)
- Grootjans A, Iturraspe R, Lanting L, Fritz C, Joosten H (2010). Ecohydrological features of some contrasting mires in Tierra del Fuego, Argentina. *Mires and Peat* 6: 01. - doi: [10.19189/001c.128351](https://doi.org/10.19189/001c.128351)
- Heinemann K, Kitzberger T, Veblen T (2000). Influences of gap microheterogeneity on the regeneration of *Nothofagus pumilio* in a xeric old-growth forest of northwestern Patagonia, Argentina. *Canadian Journal of Forest Research* 30: 25-31. - doi: [10.1139/x99-181](https://doi.org/10.1139/x99-181)
- Heinemann K, Kitzberger T (2006). Effects of position, understorey vegetation and coarse woody debris on tree regeneration in two environmentally contrasting forests of north-western Patagonia: a manipulative approach. *Journal of Biogeography* 33: 1357-1367. - doi: [10.1111/j.1365-2699.2006.01511.x](https://doi.org/10.1111/j.1365-2699.2006.01511.x)
- Henn JJ, Anderson CB, Martínez Pastur G (2016). Landscape-level impact and habitat factors associated with invasive beaver distribution in Tierra del Fuego. *Biological Invasions* 18: 1679-1688. - doi: [10.1007/s10530-016-1110-9](https://doi.org/10.1007/s10530-016-1110-9)
- Herrera-Bevan EJ, Ibáñez I (2024). Detrimental impacts of flooding conditions on native tree recruitment but not on invasive plants. *Forest Ecology and Management* 561: 121886. - doi: [10.1016/j.foreco.2024.121886](https://doi.org/10.1016/j.foreco.2024.121886)
- Hieke S, Menzel CM, Ludders P (2002). Effect of leaf, shoot and fruit development on photosynthesis of lychee trees (*Litchi chinensis*). *Tree Physiology* 22: 955-961. - doi: [10.1093/treephys/22.13.955](https://doi.org/10.1093/treephys/22.13.955)
- Hikosaka K (2021). Photosynthesis, chlorophyll fluorescence and photochemical reflectance index in photoinhibited leaves. *Functional Plant Biology* 48 (8): 815-826. - doi: [10.1071/FP20365](https://doi.org/10.1071/FP20365)
- Ivancich HS, Lencinas MV, Martínez Pastur GJ, Esteban RMS, Hernández L, Lindström I (2012). Foliar anatomical and morphological variation in *Nothofagus pumilio* seedlings under controlled irradiance and soil moisture levels. *Tree Physiology* 32 (5): 554-564. - doi: [10.1093/treephys/tps024](https://doi.org/10.1093/treephys/tps024)
- Krzeminska B, Borkowska I, Malm M, Tchorzewska D, Vangronsveld J, Vassilev A, Dos Santos Szewczyk K, Wójcik M (2024). Comparative study of the photosynthetic efficiency and leaf structure of four *Cotoneaster* species. *Scientific Reports* 14: 25113. - doi: [10.1038/s41598-024-75434-w](https://doi.org/10.1038/s41598-024-75434-w)
- Lencinas MV, Martínez Pastur GJ, Moretto A, Gallo E, Busso C (2007). Producción diferencial de biomasa en plántulas de *Nothofagus pumilio* bajo gradientes de luz y humedad del suelo [Differential biomass production in *Nothofagus pumilio* seedlings along light and soil moisture gradients]. *Bosque* 28 (3): 241-248. [in Spanish] - doi: [10.4067/s0717-92002007000300009](https://doi.org/10.4067/s0717-92002007000300009)
- Lichtenthaler HK (1987). Chlorophylls and carotenoids: pigments of photosynthetic biomembranes. *Methods in Enzymology* 148: 350-382. - doi: [10.1016/0076-6879\(87\)48036-1](https://doi.org/10.1016/0076-6879(87)48036-1)
- Lichtenthaler HK, Ač A, Marek MV, Kalina J, Urban O (2007). Differences in pigment composition, photosynthetic rates and chlorophyll fluorescence images of sun and shade leaves of four tree species. *Plant Physiology and Biochemistry* 45 (8): 577-588. - doi: [10.1016/j.plaphy.2007.04.006](https://doi.org/10.1016/j.plaphy.2007.04.006)
- Lichtenthaler HK, Babani F (2022). Contents of photosynthetic pigments and ratios of chlorophyll a/b and chlorophylls to carotenoids (a+b)/(x+c) in C_4 plants as compared to C_3 plants. *Photosynthetica* 60: 3-9. - doi: [10.32615/ps.2021.041](https://doi.org/10.32615/ps.2021.041)
- Martínez Pastur GJ, Cellini J, Peri P, Vukasovic R, Fernández C (2000). Timber production of *Nothofagus pumilio* forests by a shelterwood system in Tierra del Fuego (Argentina). *Forest Ecology and Management* 134: 153-162. - doi: [10.1016/S0378-1127\(99\)00253-4](https://doi.org/10.1016/S0378-1127(99)00253-4)
- Martínez Pastur GJ, Lencinas MV, Peri PL, Miriam Arena M (2007). Photosynthetic plasticity of *Nothofagus pumilio* seedlings to light intensity and soil moisture. *Forest Ecology and Management* 243: 274-282. - doi: [10.1016/j.foreco.2007.03.034](https://doi.org/10.1016/j.foreco.2007.03.034)

- Martínez Pastur GJ, Peri PL, Cellini JM, Lencinas MV, Barrera MD, Ivancich H (2011a). Canopy structure analysis for estimating forest regeneration dynamics and growth in *Nothofagus pumilio* forests. *Annals of Forest Science* 68: 587-594. - doi: [10.1007/s13595-011-0059-1](https://doi.org/10.1007/s13595-011-0059-1)
- Martínez Pastur GJ, Lencinas MV, Soler Esteban R, Ivancich H, Peri PL, Moretto A, Lindström I (2011b). Plasticidad ecofisiológica de plántulas de *Nothofagus pumilio* frente a combinaciones de niveles de luz y humedad en el suelo [Eco-physiological plasticity of *Nothofagus pumilio* seedlings in response to combinations of light and soil moisture levels]. *Ecología Austral* 21: 301-315. [in Spanish]
- Martínez Pastur GJ, Rodríguez-Souilla J, Bottan L, Favoretti S, Cellini JM (2024). Quantifying blowdown disturbance in overstory retention patches in managed *Nothofagus pumilio* forests with variable retention harvesting. *Forests* 15 (8): 1432. - doi: [10.3390/f15081432](https://doi.org/10.3390/f15081432)
- Mattila H, Tyystjärvi E (2023). Red pigments in autumn leaves of Norway maple do not offer significant photoprotection but coincide with stress symptoms. *Tree Physiology* 43 (5): 751-768. - doi: [10.1093/treephys/tpad010](https://doi.org/10.1093/treephys/tpad010)
- Maxwell K, Johnson GN (2000). Chlorophyll fluorescence: a practical guide. *Journal of Experimental Botany* 51 (345): 659-668. - doi: [10.1093/jexbot/51.345.659](https://doi.org/10.1093/jexbot/51.345.659)
- Mayoral C, Calama R, Sánchez-González M, Pardos M (2015). Modelling the influence of light, water and temperature on photosynthesis in young trees of mixed Mediterranean forests. *New Forests* 46 (4): 485-506. - doi: [10.1007/s11056-015-9471-y](https://doi.org/10.1007/s11056-015-9471-y)
- Mestre LM, Argañaraz CI, Preiß-Daimler I, Fernández L, Turi L, Soler R (2024). Influence of environmental conditions and initial sapling size in *Nothofagus* survival and growth: implications for restoration of burnt sub-Antarctic forests. *Austral Ecology* 49(1): e13269. - doi: [10.1111/aec.13269](https://doi.org/10.1111/aec.13269)
- Mondino V, Pastorino MJ, Gallo LA (2019). Variación altitudinal de caracteres fenológicos y crecimiento inicial en condiciones controladas entre poblaciones de *Nothofagus pumilio* provenientes del Centro-Oeste de Chubut, Argentina [Altitudinal variation in phenological traits and initial growth under controlled conditions among *Nothofagus pumilio* populations from central-western Chubut, Argentina]. *Bosque (Valdivia)* 40 (1): 87-94. [in Spanish] - doi: [10.4067/S0717-92002019000100087](https://doi.org/10.4067/S0717-92002019000100087)
- Murchie EH, Lawson T (2013). Chlorophyll fluorescence analysis: a guide to good practice and understanding some new applications. *Journal of Experimental Botany* 64 (13): 3983-3998. - doi: [10.1093/jxb/ert208](https://doi.org/10.1093/jxb/ert208)
- Murchie EH, Ruban AV (2020). Dynamic non-photochemical quenching in plants: from molecular mechanism to productivity. *The Plant Journal* 101 (4): 885-896. - doi: [10.1111/tpj.14601](https://doi.org/10.1111/tpj.14601)
- Oksanen J, Simpson GL, Blanchet FG, Kindt R, Legendre P, Minchin PR, O'Hara RB, Solymos P, Stevens MHH, Szoecs E, Wagner H, Barbour M, Bedward M, Bengtsson H, Bolker B (2025). *Vegan: community ecology package*. R package version 2:6-10. [online] URL: <http://cran.r-project.org/web/packages/vegan/vegan.pdf>
- Pérez-Harguindeguy N, Díaz S, Garnier E, Lavorel S, Poorter H, Jaureguiberry P, Bret-Harte MS, Cornwell WK, Craine JM, Gurvich DE, Urcelay C, Veneklaas EJ, Reich PB, Poorter L, Wright IJ, Ray P, Enrico L, Pausas JG, de Vos AC, Buchmann N, Funes G, Quétier F, Hodgson JG, Thompson K, Morgan HD, ter Steege H, van der Heijden MG, Sack L, Blonder B, Poschlod P, Vaieretti MV, Conti G, Staver AC, Aquino S, Cornelissen JH (2013). New handbook for standardised measurement of plant functional traits worldwide. *Australian Journal of Botany* 61 (3): 167-234. - doi: [10.1071/BT12225](https://doi.org/10.1071/BT12225)
- Peri PL, Martínez Pastur GJ, Lencinas MV (2009). Photosynthetic response to different light intensities and water status of two main *Nothofagus* species of southern Patagonian Forest, Argentina. *Journal of Forest Science* 55 (3): 101-111.
- Premoli AC, Raffaele E, Mathiasen P (2007). Morphological and phenological differences in *Nothofagus pumilio* from contrasting elevations: evidence from a common garden. *Austral Ecology* 32 (5): 515-523. - doi: [10.1111/j.1442-9993.2007.01720.x](https://doi.org/10.1111/j.1442-9993.2007.01720.x)
- R Core Team (2018). R: a language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. [online] URL: <http://www.r-project.org>
- Rebertus A, Veblen T (1993). Structure and tree-fall gap dynamics of old-growth *Nothofagus* forests in Tierra del Fuego, Argentina. *Journal of Vegetation Science* 4 (5): 641-654. - doi: [10.2307/3236129](https://doi.org/10.2307/3236129)
- Rodríguez-Souilla J, Cellini JM, Lencinas MV, Rogí FA, Chaves JE, Acuña MCA, Peri PL, Martínez Pastur GJ (2023). Variable retention harvesting and climate variations influence over natural regeneration dynamics in *Nothofagus pumilio* forests of Southern Patagonia. *Forest Ecology and Management* 544: 121221. - doi: [10.1016/j.for eco.2023.121221](https://doi.org/10.1016/j.for eco.2023.121221)
- Rojas-Rioseco M, Castillo RD, González-Campos J, Ipinza R, Sanhueza MI, Hasbún R (2023). Phylogeographic origin authentication of *Araucaria araucana* (Mol.) K Koch seedlings through the application of spectroscopy techniques in different infrared ranges and chemometric methods. *New Forests* 54 (3): 467-489. - doi: [10.1007/s11056-022-09933-x](https://doi.org/10.1007/s11056-022-09933-x)
- Sims DA, Gamon JA (2002). Relationships between leaf pigment content and spectral reflectance across a wide range of species, leaf structures and developmental stages. *Remote Sensing Environment* 81: 337-354. - doi: [10.1016/S0034-4257\(02\)00010-X](https://doi.org/10.1016/S0034-4257(02)00010-X)
- Taiz L, Møller IM, Murphy A, Zeiger E (2023). *Plant physiology and development* (7th edn). Oxford University Press, Oxford, UK, pp. 572.
- Takahashi S, Murata N (2008). How do environmental stresses accelerate photoinhibition? *Trends in Plant Science* 13 (4): 178-182. - doi: [10.1016/j.tplants.2008.01.005](https://doi.org/10.1016/j.tplants.2008.01.005)
- Tian YN, Zhong RH, Wei JB, Luo HH, Eyal Y, Jin HL, Wu LJ, Liang KY, Li YM, Chen SZ, Zhang ZQ, Pang XQ (2021). *Arabidopsis* CHLOROPHYLLASE 1 protects young leaves from long-term photodamage by facilitating FtsH-mediated D1 degradation in photosystem II repair. *Molecular Plant* 14 (7): 1149-1167. - doi: [10.1016/j.molp.2021.04.006](https://doi.org/10.1016/j.molp.2021.04.006)
- Tognetti R, Minotta G, Pinzauti S, Michelozzi M, Borghetti M (1998). Acclimation to changing light conditions of long-term shade-grown beech (*Fagus sylvatica*) seedlings of different geographic origins. *Trees* 12: 326-333. - doi: [10.1007/PL00009719](https://doi.org/10.1007/PL00009719)
- Toro-Manríquez M, Cellini JM, Lencinas MV, Peri PL, Peña Rojas KA, Martínez Pastur GJ (2019). Suitable conditions for natural regeneration in variable retention harvesting of southern Patagonian *Nothofagus pumilio* forests. *Ecological Processes* 8: 18. - doi: [10.1186/s13717-019-0175-7](https://doi.org/10.1186/s13717-019-0175-7)
- Torres Netto A, Campostrini E, De Oliveira JG, Bressan-Smith RE (2005). Photosynthetic pigments, nitrogen, chlorophyll a fluorescence and SPAD-502 readings in coffee leaves. *Scientia Horticulturae* 104 (2): 199-209. - doi: [10.1016/j.scientia.2004.08.013](https://doi.org/10.1016/j.scientia.2004.08.013)
- Van Galen LG, Lord JM, Orlovich DA, Larcombe MJ (2021). Restoration of southern hemisphere beech (*Nothofagaceae*) forests: a meta-analysis. *Restoration Ecology* 29(3): e13333. - doi: [10.1111/rec.13333](https://doi.org/10.1111/rec.13333)
- Voesenek LA, Bailey-Serres J (2015). Flood adaptive traits and processes: an overview. *New Phytologist* 206 (1): 57-73. - doi: [10.1111/nph.13209](https://doi.org/10.1111/nph.13209)
- Yang XL, Ye ZWY, Kang HJ, Robakowski P, Ye ZP, Wang FB, Zhou SX (2025). Modeling light response of effective quantum efficiency of photosystem II for C₃ and C₄ crops. *Frontiers in Plant Science* 16: 1478346. - doi: [10.3389/fpls.2025.1478346](https://doi.org/10.3389/fpls.2025.1478346)
- Zait Y, Shemer OE, Cochavi A (2024). Dynamic responses of chlorophyll fluorescence parameters to drought across diverse plant families. *Physiologia Plantarum* 176: e14527. - doi: [10.1111/ppl.14527](https://doi.org/10.1111/ppl.14527)
- Zhang Y, Chen X, Geng S, Zhang X (2025). A review of soil waterlogging impacts, mechanisms, and adaptive strategies. *Frontiers in Plant Science* 16: 1545912. - doi: [10.3389/fpls.2025.1545912](https://doi.org/10.3389/fpls.2025.1545912)

Supplementary Material

Fig. S1 - Seasonal variation in air relative humidity, air temperature, and soil temperature during the growing season under three light-intensity treatments (high, medium, and low) in the greenhouse.

Tab. S1 - ANOVA results of fluorescence parameters in *Nothofagus pumilio* seedlings, considering month, light intensity, and soil moisture as factors and their interactions.

Fig. S2 - Pearson's correlation coefficients between all response variables of *Nothofagus pumilio* seedlings.

Tab. S2 - Loadings of eco-physiological variables on the first two principal components (PC1 and PC2) from the PCA performed for October, January, and March.

Link: Bottan_5020@suppl001.pdf