

Spruce budworm biological and nutritional performance responses to varying levels of monoterpenes

Meriç Kumbasli⁽¹⁾, Éric Bauce⁽²⁾

Dose effect of six monoterpenes (α -pinene, bornyl acetate, camphene, δ -3-carene, terpinolene, tricyclene) found in the foliage of host trees was tested on sixth-instar spruce budworm (*Choristoneura fumiferana* Clem.) using artificial diet. The larval mortality, growth and food utilization have been observed. Two monoterpenes, α -pinene and δ -3-carene caused 22 and 12% mortality respectively at concentrations found in balsam fir foliage. Bornyl acetate and camphene reduced larval survival when their concentration was higher than the foliage. Terpinolene and tricyclene have no effect on mortality. All six tested monoterpenes reduced larval growth rate. Spruce budworm tried to minimize this negative growth impact by increasing his digestibility in presence of camphene and δ -3-carene, and by increasing his efficiency of conversion of ingested-digested food with α -pinene and bornyl acetate. These results support the traditional theory that monoterpenes are a defense agent against spruce budworm and that each monoterpene has a different mode of action and effects which are not necessarily proportional to its concentration.

Keywords: Herbivore Defense, Monoterpenes, Insect-plant Interaction

Introduction

Spruce budworm (*Choristoneura fumiferana* (Clem.)) is an important defoliator of coniferous forests in eastern North America that can severely damage balsam fir (*Abies balsamea* (L.) Mill.) and three spruce tree species (*Picea glauca* (Moench) Voss, *P. mariana* (Mill.) BSP, *P. rubens* Sarg. - Sanders 1991). Monoterpenes are among the foliar secondary metabolites deployed by these host plant species to defend against spruce budworm. Concentrations of these compounds, like others present in the host plant foliage, are subject to spatial and temporal change. It is well known that terpenoid

concentrations in foliar tissues generally increase with light intensity (Spring et al. 1986, Bryant et al. 1987, Gref & Tenow 1987), with nutrient availability (Björkman et al. 1998), and following stand thinning, for example, in balsam fir (Bauce 1996, Lamontagne et al. 2000, Fuentealba & Bauce 2012).

Bauce et al. (1994) showed that monoterpenes are positively correlated with the strong resistance offered by young balsam fir trees against budworm attack, compared to mature individuals. Kumbasli et al. (2011) simulated this phenomenon under laboratory conditions using an artificial diet. They found that high monoterpenes concentrations that reflected the levels of monoterpenes found in young balsam fir trees increased spruce budworm mortality, while decreasing spruce budworm growth rate and efficiency of food conversion. The authors used a mixture of ten synthetic monoterpenes that represented levels of foliage monoterpenes (per dry mass), which were encountered in young and old balsam fir trees. While the defensive role of this monoterpene mixture was demonstrated in this study, but it did not provide information regarding the effects of each monoterpene found in the foliage or its most effective concentration. The mode of action varied, depending on the molecule and also its concentration. For example, Cates et al. (1987) measured dosage effects of bornyl acetate and β -pinene on the western spruce budworm (*Choristoneura occi-*

dentalis). These authors reported that bornyl acetate reduced both growth and survival, β -pinene increased budworm growth rates by functioning as a feeding stimulant. All these studies show that spruce budworm is affected in its feeding behavior by monoterpenes, but there is little information on the individual effect of these compounds on the nutritional indices and mortality.

We performed an experiment under laboratory conditions using an artificial diet to determine the individual dosage effects of six monoterpenes (α -pinene, bornyl acetate, camphene, δ -3-carene, terpinolene, tricyclene), which were naturally found in the host tree foliage, on food utilization and survival of sixth-instar spruce budworm larvae.

Materials and methods

The principal monoterpenes that are produced by host tree foliage and their concentrations have been determined in several previous studies (Bauce et al. 1994, Bauce 1996, Carisey & Bauce 1997a, 1997b, 1997c, Kumbasli et al. 2011). Based on these data, six monoterpenes (α -pinene, bornyl acetate, camphene, δ -3-carene, terpinolene, tricyclene - Sigma-Aldrich, Milwaukee, WI, USA) were tested at different concentrations (Tab. 1). Two different levels were selected for each monoterpene in order to test the effect of concentrations within and exceeding their natural range in the foliar tissue. The exact concentrations were determined based on the preliminary study, concentration ranges, and methodology of the laboratory study (*i.e.*, incorporation of monoterpene to the artificial diet). Artificial diet was prepared as described by McMorran (1965) and each monoterpene concentration was incorporated into the diet by hexane solubilization. A diet without monoterpenes was also prepared and considered as the control for each monoterpene molecule. The artificial diet was poured into 37 mL cups (Solo Cup Company, Chicago, IL, USA) and five additional cups were used to determine the dry mass of food provided to the larvae. Each diet cup was weighed before and after feeding, and the difference was multiplied by the food dry mass ratio to determine the quantity of dry food that had been ingested. We focused upon sixth-instar larvae to understand the effects of monoterpenes because 87% of spruce budworm total food consumption occurs during this larval stage (Miller 1977). Further, Carisey & Bauce (1997b) found that foliar monoterpene concentrations had no effect on nutritional indices calculated for fifth-instar larvae. The monoterpenes used were in a volatile form, with preliminary tests showing that 7 days was a sufficient period of time for deploying the diets without marked decreases in their monoterpene concentrations. Spruce bud-

□ (1) Faculty of Forestry, Istanbul University, 34473 Bahçeköy, Istanbul (Turkey); (2) Département des Sciences du Bois et de la Forêt, Faculté de Foresterie et de Géomatique, Université Laval, G1V 0A6 Québec, QC (Canada)

@ Meriç Kumbasli
(kumbasli@istanbul.edu.tr)

Received: Jan 24, 2013 - Accepted: Apr 09, 2013

Citation: Kumbasli M, Bauce E, 2013. Spruce budworm biological and nutritional performance responses to varying levels of monoterpenes. iForest 6: 310-314 [online 2013-07-16] URL: <http://www.sisef.it/forest/contents/?id=ifor0956-006>

Communicated by: Massimo Faccoli

worm larvae were obtained from the rearing facility of the Great Lakes Forestry Centre, Canadian Forest Service (Sault Ste. Marie, Ontario, Canada). Twenty-five sixth-instar females were individually reared in each monoterpene assay, with larvae being monitored twice a day to record mortality and nutritional indices measurements (initial and final larval mass, ingested food, faeces excreted) over 7 days (duration that covers the last instar of spruce budworm). Insect rearing conditions were maintained at 23 °C, 65% relative humidity, and a 16 h: 8 h LD photoperiod. Nutritional indices were expressed on a dry-mass basis according to Waldbauer (1968), and included (eqn. 1 to 5):

$$RGR = \frac{G}{MW \cdot \text{development time}(h)}$$

$$RCR = \frac{I}{MW \cdot \text{development time}(h)}$$

$$D = \frac{I-F}{I} \cdot 100$$

$$ECI = \frac{G}{I} \cdot 100$$

$$ECD = \frac{G}{I-F} \cdot 100$$

where *RGR* is the relative growth rate, *RCR* is the relative consumption rate, *D* is the digestibility, *ECI* is the efficiency of conversion of ingested food, *ECD* is the efficiency of conversion of digested food, *G* is the mass gained (= final mass - initial mass), *MW* is the mean larval mass [= *G* / log(final mass/initial mass)], *I* is the ingested food, *F* is the faeces.

Nutritional indices were analyzed by a general linear model procedure, *i.e.*, a covariance analysis described by Bauce et al. (1994). Because *RCR* and *RGR* indices are ratios and the variance of a ratio is equal to the variances only when the dominator of the ratio is constant, they were adjusted in accordance with Bauce et al. (1994). Mortality was treated in a binomial analysis using the GENMOD procedure in SAS (SAS Institute 2003), followed by LSMEANS to detect differences between concentrations.

Results

The terpenes α -pinene and δ -3-carene exerted significant mortality effects on sixth-instar budworm larvae. These effects were most pronounced at the maximum concentrations found in the foliage (Fig. 1, Tab. 1).

Tab. 1 - Monoterpenes concentrations measured in *A. balsamea* foliage and doses used in the experiment. (a) sources: Bauce et al. 1994, Bauce 1996, Carisey & Bauce 1997a, 1997b, 1997c.

Monoterpenes	% dry weight	
	Min and max % observed in foliage (a)	Doses used
α -pinene	0.05 - 0.71	0 - 0.6 - 1 - 3
Bornyl acetate	0.019 - 0.49	0 - 0.3 - 0.8 - 2.5
Camphene	0.011 - 0.33	0 - 0.2 - 0.5
δ -3-carene	0 - 0.58	0 - 0.2 - 0.5 - 1.5
Terpinolene	0 - 0.046	0 - 0.03 - 0.05 - 0.15
Tricyclene	0.003 - 0.057	0 - 0.04 - 0.09 - 0.3

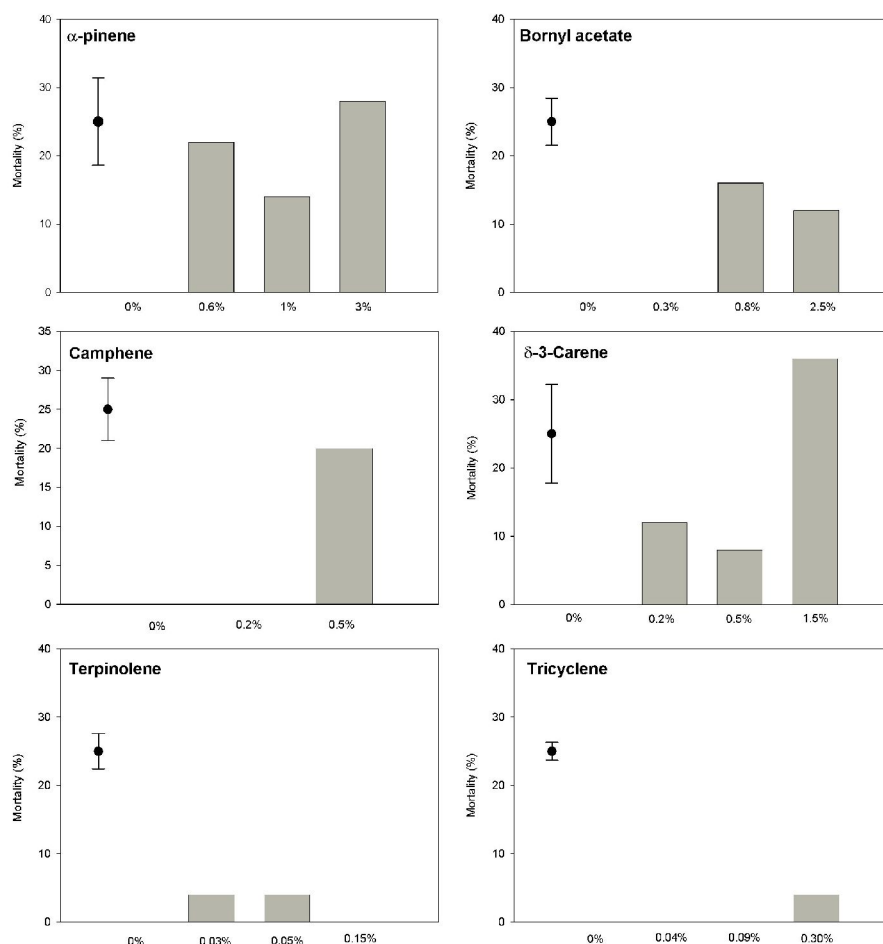


Fig. 1 - Mortality after seven days of female sixth-instar spruce budworm fed on artificial diet supplemented different levels and different monoterpenes. Bar represents ± 2 SE (*n* = 25 larvae per treatment).

Tab. 2 - The effect of monoterpene concentrations on *C. fumiferana* nutritional indices. Insects were reared on artificial diet supplemented with synthetic monoterpenes. Data were analyzed using ANCOVA. (*): significant effect (*P* < 0.01); (ns): no significant effect.

Index	α -pinene		Bornyl acetate		Camphene		δ -3-carene		Terpinolene		Tricyclene	
	F _(3; 83)	P	F _(3; 89)	P	F _(2; 65)	P	F _(3; 80)	P	F _(3; 94)	P	F _(3; 94)	P
<i>D</i>	101.83	0.0001*	1.92	0.0131 ^{ns}	7.88	0.0009*	20.31	0.0001*	1.95	0.1270 ^{ns}	2.63	0.0549 ^{ns}
<i>ECD</i>	175.18	0.0001*	3.37	0.0221 ^{ns}	0.33	0.7196 ^{ns}	27.12	0.0001*	2.89	0.0394 ^{ns}	3.65	0.0153 ^{ns}
<i>ECI</i>	68.53	0.0001*	5.61	0.0014*	0.26	0.7735 ^{ns}	19.26	0.0001*	1.29	0.2809 ^{ns}	1.13	0.3411 ^{ns}
<i>RCR</i>	31.82	0.0001*	6.47	0.0005*	0.91	0.4070 ^{ns}	42.86	0.0001*	2.46	0.0678 ^{ns}	0.28	0.8417 ^{ns}
<i>RGR</i>	173.18	0.0001*	5.63	0.0014*	3.57	0.0340 ^{ns}	146.38	0.0001*	6.53	0.0005*	2.83	0.0427 ^{ns}

Tab. 3 - Nutritional indices (mean \pm standard deviation) of sixth-instar female spruce budworm larvae fed on artificial diet containing different concentrations of monoterpenes. Values in each column followed by the same letter do not differ significantly at $P < 0.05$ according Duncan's multiple range test.

Monoterpene	Concentration (%)	Nutritional Indices				
		D (%)	ECD (%)	ECI (%)	RCR (mg mg ⁻¹ h ⁻¹ 10 ⁻²)	RGR (mg mg ⁻¹ h ⁻¹ 10 ⁻³)
α -pinene	0	48.30 \pm 0.35 ^a	50.11 \pm 1.15 ^a	23.27 \pm 0.50 ^a	6.07 \pm 0.12 ^a	13.96 \pm 0.19 ^a
	0.6	38.69 \pm 0.50 ^b	79.99 \pm 1.43 ^b	28.84 \pm 0.69 ^b	4.24 \pm 0.13 ^b	12.80 \pm 0.25 ^b
	1	44.76 \pm 0.68 ^c	62.61 \pm 1.42 ^c	28.49 \pm 0.71 ^b	4.40 \pm 0.13 ^b	12.42 \pm 0.20 ^b
	3	63.36 \pm 0.79 ^d	21.73 \pm 1.31 ^d	14.81 \pm 0.61 ^c	4.26 \pm 0.18 ^b	6.53 \pm 0.35 ^c
Bornyl acetate	0	44.73 \pm 0.40	51.02 \pm 1.46	23.09 \pm 0.67	5.52 \pm 0.21	12.44 \pm 0.15
	0.3	44.06 \pm 0.40	54.26 \pm 1.34	23.37 \pm 0.57	5.24 \pm 0.14	12.15 \pm 0.20
	0.8	46.00 \pm 0.45	52.94 \pm 1.42	24.06 \pm 0.72	4.98 \pm 0.22	12.75 \pm 0.24
	2.5	45.16 \pm 0.81	57.27 \pm 1.46	26.56 \pm 0.68	4.39 \pm 0.16	11.58 \pm 0.21
Camphene	0	41.57 \pm 0.56	58.06 \pm 1.29	24.40 \pm 0.67	5.57 \pm 0.19	13.83 \pm 0.17
	0.2	43.67 \pm 0.48	56.90 \pm 1.20	24.86 \pm 0.57	5.52 \pm 0.15	13.62 \pm 0.16
	0.5	44.66 \pm 0.63	56.81 \pm 1.08	24.21 \pm 0.74	5.26 \pm 0.15	13.11 \pm 0.23
δ -3-carene	0	43.42 \pm 0.47 ^a	55.40 \pm 1.19 ^a	24.43 \pm 0.56 ^a	5.18 \pm 0.21 ^a	12.45 \pm 0.22 ^a
	0.2	50.14 \pm 0.85 ^b	49.49 \pm 0.97 ^b	24.95 \pm 0.50 ^a	4.88 \pm 0.15 ^a	12.07 \pm 0.15 ^a
	0.5	51.81 \pm 0.83 ^b	48.18 \pm 1.09 ^b	24.50 \pm 0.59 ^a	4.17 \pm 0.23 ^b	10.07 \pm 0.20 ^b
	1.5	47.01 \pm 1.32 ^c	38.01 \pm 2.01 ^c	18.09 \pm 1.12 ^b	1.97 \pm 0.17 ^c	6.50 \pm 0.24 ^c
Terpinolene	0	40.99 \pm 0.49	62.27 \pm 1.06	24.93 \pm 0.50	5.30 \pm 0.19	13.09 \pm 0.25 ^a
	0.03	43.26 \pm 0.89	59.81 \pm 1.05	25.88 \pm 0.58	5.22 \pm 0.22	13.04 \pm 0.29 ^a
	0.05	41.47 \pm 0.79	58.39 \pm 1.32	24.77 \pm 0.65	5.44 \pm 0.17	13.29 \pm 0.21 ^a
	0.15	42.25 \pm 0.59	62.86 \pm 1.43	26.21 \pm 0.71	4.69 \pm 0.23	11.87 \pm 0.24 ^b
Tricyclene	0	42.32 \pm 0.66	58.14 \pm 1.02	24.59 \pm 0.55	5.09 \pm 0.17	13.49 \pm 0.24
	0.04	44.27 \pm 0.70	56.80 \pm 1.13	25.06 \pm 0.56	5.00 \pm 0.10	13.44 \pm 0.19
	0.09	44.21 \pm 0.47	56.52 \pm 0.92	24.98 \pm 0.50	5.18 \pm 0.15	13.89 \pm 0.17
	0.3	44.47 \pm 0.60	53.35 \pm 1.10	23.72 \pm 0.66	5.13 \pm 0.13	13.02 \pm 0.22

Bornyl acetate and camphene had significant effects when their concentrations were twice those encountered in the foliage. Any effect from terpinolene and tricyclene on mortality was not observed. The greatest mortality (22%) was observed for α -pinene at a concentration of 0.6%. A slightly lower mortality level was detected for δ -3-carene. All larvae in the control treatments were alive after 7 days, compared to those exposed to each of the monoterpenes (Fig. 1).

Results of ANCOVA performed indicated that α -pinene, bornyl acetate, camphene, δ -3-carene and terpinolene significantly affected sixth-instar spruce budworm nutritional indices (Tab. 2). The presence of monoterpenes, except for bornyl acetate, decreased RGR. Digestibility increased with increasing concentrations of camphene and δ -3-carene when compared to the control; decreased digestibility was only observed for α -pinene. ECD and ECI indices increased with increased concentrations of α -pinene and bornyl acetate. ECD decreased in the presence of δ -3-carene. Nutritional indices of sixth-instar spruce budworm that were reared on artificial diets containing different concentrations of monoterpenes are summarized in Tab. 3.

Discussion

The defensive role of monoterpenes against spruce budworm has been demonstrated in several studies, based on rearing trials either on foliage (Bauce et al. 1994) or on artificial

diet (Kumbasli et al. 2011). This negative response was supported by results of the present study, given that we detected increased mortality (α -pinene, δ -3-carene) and reduced growth rates (α -pinene, δ -3-carene, terpinolene) as result of exposure to the individual monoterpenes. In addition, this study allowed us to reveal the mode of action for each monoterpene as previous studies used only mixtures of monoterpenes.

Secondary compounds can negatively affect herbivores performance in different ways: (i) by reducing food intake; and once ingested (ii) by reducing the efficiency of food utilization; and (iii) through their direct toxic effects (Elsayed 2011). Mortality is frequently used to assess the effect of secondary compounds on insect biological performance and the toxicity of secondary compounds is often the main argument advanced for accepting the defensive functions of secondary compounds in plant-herbivore interactions. The higher spruce budworm mortality observed in young balsam fir compared to old trees was attributed to high concentrations of δ -3-carene, terpinolene and bornyl acetate (Bauce et al. 1994, Kumbasli et al. 2011). Similarly, Bauce & Kumbasli (2007) found that only resistant white spruce trees (*Picea glauca* Moench) contain borneol and δ -3-carene. Our results show that α -pinene and δ -3-carene individually provide good protection against herbivory at concentrations that were encountered in the foliage.

Exceeding the maximum level found in the foliage does not necessarily increase spruce budworm mortality, especially in the case of α -pinene. We observed mortality levels that were three times higher for δ -3-carene when its concentrations were seven times higher than the maximum routinely found in foliage (from 0.2 to 1.5%). Bornyl acetate and camphene must reach a concentration that is two-fold higher than the foliage to achieve a toxic effect.

In the literature (Städler 1974, Chen et al. 2002, Grant et al. 2007), α -pinene has been reported as budworm oviposition stimulant, but our results showed that this monoterpene, at concentrations found in the foliage, proved toxic to sixth-instar larvae, incurring 20% mortality. To reduce the toxic effect of α -pinene, spruce budworm decreases the digestibility *D* (Tab. 3), i.e., it attempts to minimize the amount of α -pinene retained in its body. An opposite strategy is employed for δ -3-carene. To reduce negative effects of δ -3-carene, spruce budworm larvae decrease their RCR by increasing the rate of food transit through the gut.

Bornyl acetate, δ -3-carene, camphene and terpinolene are considered as being toxic to spruce budworm (Mattson et al. 1991, Bauce et al. 1994, Kumbasli et al. 2011). Our results show that these monoterpenes are individually not very effective in terms of their toxicity against sixth-instar larvae. This low toxicity can be explained by the absence of

other monoterpenes or by the provision of sufficient energy for detoxification by soluble sugars.

Our results on nutritional indices show that the reaction of spruce budworm differed for each monoterpene against which it was tested. We observed a small decrease of *RGR* in the presence of α -pinene, bornyl acetate and δ -3-carene, while other monoterpenes do not affect growth. To counteract the negative effects of δ -3-carene, the spruce budworm tries to increase its digestibility. Despite this attempt, detoxification of δ -3-carene is an energetically costly process, which results in a reduced efficiency of conversion (typical compensation strategy observed with tannins). This compensation strategy is not validated in the case of α -pinene, where digestibility is reduced compared to the diet without the monoterpene and a correspondingly higher efficiency of conversion. A reduced digestibility compared to control diet can be attributed to the phago-repulsive effects of α -pinene. All of these different strategies used by spruce budworm to reduce negative effects of monoterpenes observed in this study can explain why these molecules are more effective as an ensemble in terms of foliage protection. Using a single strategy is not sufficient to confront and counteract the other monoterpenes present in the foliage.

Carisey & Bauce (1997a) tried to explain the lower efficiency of food conversion and reduced *RGR* of spruce budworm that were fed on old balsam fir foliage, which they attributed to a lack of water, to nutritional unbalances, and to a high level of some monoterpenes such as bornyl acetate, terpinolene and camphene. Our results showed that these three monoterpenes, when tested individually, reduced neither growth nor the efficiency of food conversion. This demonstrates that the reductions observed in the work of Carisey & Bauce (1997a) were probably due to a lack of water or nutritional imbalance within the old foliage.

We know that factors such as nitrogen or soluble sugar levels may be important in the compensatory mechanism employed by insects to deal with secondary compounds. Cates et al. (1987) emphasizes the importance of nitrogen concentration on the effect of bornyl acetate and β -pinene. In our study, nitrogen and soluble sugars were maintained at stable levels to assess the effect of each secondary compound. Soluble sugars play an important role during the sixth-instar stage of spruce budworm and it would be interesting to observe the effects of these monoterpenes when combined with different concentrations of soluble sugars.

In conclusion, the negative effects of monoterpenes on spruce budworm performance and the respective variations of in their modes of action were demonstrated in the

current study. It has been widely assumed that the effect of secondary compounds on herbivores is proportional to concentrations of the compounds of interest (Mattson et al. 1991). Our results do not support this assumption and we can conclude that spruce budworm host plants produce these molecules at concentrations that provide optimal protection.

Acknowledgements

The authors wish to thank Sophie Rochefort, Martin Charest and Richard Bérubé (Université Laval, Quebec, Canada) for their helpful contributions to this study and Dr. W.F.J. Parsons for checking the English. Financial support was provided by the Natural Sciences and Engineering Research Council of Canada (NSERC), the *Ministère des Ressources Naturelles et de la Faune du Québec* (MRNFQ), and the Scientific and Technological Research Council of Turkey (TÜBİTAK) BİDEB-2219 Postdoctoral Research Program. We declare that the experiments comply with the current laws of the country in which they were performed.

References

- Bauce E, Crépin M, Carisey N (1994). Spruce budworm growth, development and food utilization on young and old balsam fir trees. *Oecologia* 97: 499-507. - doi: [10.1007/BF00325888](https://doi.org/10.1007/BF00325888)
- Bauce E (1996). One and 2 years impact of commercial thinning on spruce budworm feeding ecology and host tree foliage production and chemistry. *Forestry Chronicle* 72 (4): 393-398.
- Bauce E, Kumbasli M (2007). Natural resistance of fast growing white spruce, *Picea glauca* (Moench), trees against spruce budworm, *Choristoneura fumiferana* (Clem.). In: Proceeding of the International Symposium "Bottlenecks, Solutions, and Priorities in the Context of Functions of Forest Resources" (Demir M, Yilmaz E eds). Istanbul (Turkey) 17-19 October 2007, pp. 687-695.
- Björkman C, Kyto M, Larsson S, Niemela P (1998). Different responses of two carbon-based defences in Scots pine needles to nitrogen fertilization. *Écoscience* 5 (4): 502-507.
- Bryant JP, Chapin III FS, Reichardt PB, Clausen TP (1987). Response of winter chemical defense in Alaskan paper birch and green alder to manipulation of plant carbon/nutrient balance. *Oecologia* 72: 510-514. - doi: [10.1007/BF00378975](https://doi.org/10.1007/BF00378975)
- Carisey N, Bauce É (1997a). Impact of balsam fir foliage age on sixth-instar spruce budworm growth, development and food utilization. *Canadian Journal of Forest Research* 27: 257-264. - doi: [10.1139/x97-002](https://doi.org/10.1139/x97-002)
- Carisey N, Bauce É (1997b). Impact of balsam fir flowering on pollen and foliage biochemistry in relation to spruce budworm growth, development and food utilization. *Entomologia Experimentalis et Applicata* 85: 17-31. - doi: [10.1046/j.1570-7458.1997.00231.x](https://doi.org/10.1046/j.1570-7458.1997.00231.x)
- Carisey N, Bauce É (1997c). Balsam fir foliar

chemistry in middle and lower crowns and spruce budworm growth, development, food and nitrogen utilization. *Journal of Chemical Ecology* 23: 1963-1978. - doi: [10.1023/B:JOEC.0000006483.52480.c4](https://doi.org/10.1023/B:JOEC.0000006483.52480.c4)

Cates RG, Henderson CB, Redak RA (1987). Responses of the western spruce budworm to varying levels of nitrogen and terpenes. *Oecologia* 73: 312-316. - doi: [10.1007/BF00377524](https://doi.org/10.1007/BF00377524)

Chen Z, Kolb TE, Clancy KM (2002). The role of monoterpenes in resistance of Douglas-fir to western spruce budworm defoliation. *Journal of Chemical Ecology* 28 (5): 897-920. - doi: [10.1023/A:1015297315104](https://doi.org/10.1023/A:1015297315104)

Elsayed G (2011). Plant secondary substances and insects behaviour. *Archives of Phytopathology and Plant Protection* 44: 1534-1549. - doi: [10.1080/03235408.2010.507957](https://doi.org/10.1080/03235408.2010.507957)

Fuentealba A, Bauce É (2012). Site factors and management influence short-term host resistance to spruce budworm, *Choristoneura fumiferana* (Clem.), in a species-specific manner. *Pest Management Science* 68: 245-253. - doi: [10.1002/ps.2253](https://doi.org/10.1002/ps.2253)

Grant GG, Guo J, MacDonald L, Coppens MD (2007). Oviposition response of spruce budworm (Lepidoptera: Tortricidae) to host terpenes and green volatiles. *Canadian Entomologist* 139 (4): 564-575. - doi: [10.4039/n06-079](https://doi.org/10.4039/n06-079)

Gref R, Tenow O (1987). Resin acid variation in sun and shade needles of Scots pine (*Pinus sylvestris* L.). *Canadian Journal of Forest Research* 17: 346-349. - doi: [10.1139/x87-058](https://doi.org/10.1139/x87-058)

Kumbasli M, Bauce É, Rochefort S, Crépin M (2011). Effects of tree age and stand thinning related variations in balsam fir secondary compounds on spruce budworm *Choristoneura fumiferana* development, growth and food utilization. *Agricultural and Forest Entomology* 13: 131-141. - doi: [10.1111/j.1461-9563.2010.00505.x](https://doi.org/10.1111/j.1461-9563.2010.00505.x)

Lamontagne M, Margolis H, Bauce É (2000). Testing the ecophysiological basis for the control of monoterpene concentrations along canopy profiles in thinned and unthinned balsam fir stands. *Oecologia* 124: 318-331. - doi: [10.1007/s004420000393](https://doi.org/10.1007/s004420000393)

Mattson WJ, Haack RA, Lawrence RK, Slocum SS (1991). Considering the nutritional ecology of the spruce budworm in its management. *Forest Ecology and Management* 39: 183-210. - doi: [10.1016/0378-1127\(91\)90176-V](https://doi.org/10.1016/0378-1127(91)90176-V)

McMorran A (1965). A synthetic diet for the spruce budworm *Choristoneura fumiferana* (Clem.) (Lepidoptera: Tortricidae). *Canadian Entomologist* 97: 58-62. - doi: [10.4039/Ent9758-1](https://doi.org/10.4039/Ent9758-1)

Miller CA (1977). The feeding impact of spruce budworm on balsam fir. *Canadian Journal of Forest Research* 7: 76-84. - doi: [10.1139/x77-011](https://doi.org/10.1139/x77-011)

Sanders CJ (1991). Biology of North American spruce budworms. In: "Tortricid Pests, their Biology, Natural Enemies and Control" (Van der Geest LPS, Evenhuis HH eds). Elsevier Science Publishers BV, The Netherlands, pp. 579-620.

SAS Institute (2003). SAS/STAT User's Guide,

release 9.1. SAS Institute Inc., Cary, NC, USA.

Spring O, Priester T, Hager A (1986). Light-induced accumulation of sesquiterpene lactones in sunflower seedlings. *Journal of Plant Physiology* 123: 79-89. - doi: [10.1016/S0176-1617\(86\)](https://doi.org/10.1016/S0176-1617(86)80068-2)

[80068-2](https://doi.org/10.1016/S0176-1617(86)80068-2)

Städler E (1974). Host plant stimuli affecting oviposition behaviour of the Eastern spruce budworm. *Entomologia Experimentalis et Applicata* 17(2): 176-188. - doi: [10.1111/j.1570-7458.](https://doi.org/10.1111/j.1570-7458.1974.tb00334.x)

[1974.tb00334.x](https://doi.org/10.1111/j.1570-7458.1974.tb00334.x)

Waldbauer GP (1968). The consumption and utilization of food by insects. *Advances in Insect Physiology* 5: 229-288. - doi: [10.1016/S0065-2806\(08\)60230-1](https://doi.org/10.1016/S0065-2806(08)60230-1)